

Soluble Epoxide Hydrolase Inhibitors: New Molecules with Potential for Use in Veterinary Medicine

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ABSTRACT

Treatments for inflammation and pain are important consideration in human and veterinary medicine. The classical drugs for treatments of inflammation and pain act by inhibition of cyclooxygenase and lipoxygenase pathways. However, there is still a need to develop new veterinary drugs and trials to apply human drugs to the veterinary field in order to increase the veterinary drug armamentarium. However, it is pivotal to experimentally test human drugs and therapies in veterinary species before veterinary clinical applications. The soluble epoxide hydrolase inhibitors (sEHIs), are novel active ingredients shown to have a number of beneficial effects. This has been especially demonstrated in many animal models in relation to inflammation and pain. The present review reports the state of the art of soluble epoxide hydrolase inhibitors and suggests their potential use in veterinary medicine.

Keywords: Soluble epoxide hydrolase, epoxyeicosatrienoic acids, inflammation, pain.

INTRODUCTION

Pets are treated as members of the family and pet owners demand the same level of care they expect for themselves. This change in attitude has led to a rapid evolution in the field of pharmacology with a trend towards the development of more effective and innovative veterinary therapies with higher potency, more rapid speed of action and fewer side effects (1).

Treatment of pain and inflammation are important considerations in human medicine. Likewise, in veterinary medicine in recent years, pain has been shown to dramatically affect animal welfare and production, and interest in the field of analgesia is increasing (2). Furthermore, veterinary pharmacology still has a limited drug armamentarium and human drugs are increasingly being investigated for veterinary use. It has only been in recent years that analgesics have been marketed exclusively for veterinary patients. Therefore, it is pivotal that new human drugs and therapies be tested also in veterinary species (1).

The two main classes of drugs used to reduce pain in animals are opioids and nonsteroidal anti-inflammatory drugs (NSAIDs). Recently, some of the novel molecules in these classes marketed for the human field have been successfully tested on veterinary species (3-5). In the last few years, many researchers have directed attention towards arachidonic acid (AA) metabolism and in particular, to the cytochrome P450 (CYP450) enzymes. These have been referred to as the third pathway of AA metabolism, in addition to cyclooxygenases (COX) and lipoxygenases (LOX) (6).

All AA metabolites, which encompass the prostanoids, leukotrienes and epoxy fatty acids (Figure 1), are bioactive lipids that play a positive or negative role in inflammation and pain, specifically under pathological conditions. The allogeic and pro-inflammatory prostanoids and leukotrienes drive and maintain inflammation, while the anti-inflammatory and analgesic epoxy fatty acids epoxyeicosatrienoic acids (EETs) reduce and resolve inflammation (7).

Compared to the well-recognized products of the COX and LOX branches of the AA cascade, the EETs generated by CYPs were only discovered in the early 1980s (7). EETs have various beneficial biological effects: not only do they have anti-inflammatory and analgesic actions but they also have protective effects on the cardiovascular system and kidney as recently reported in the literature (8-19) (Table 1).

Table 1: Functions of EETs

| Functions | References |
|---|------------|
| Effect on sodium ion channel | 8 |
| Increases in intracellular Ca ²⁺ | 9 |
| Activation of K _{ATP} ion channel | 10 |
| Ca ²⁺ signalling | 11, 12 |
| Activation of BK _{Ca} ion channel | 13 |
| Myocardocyte contraction | 14 |
| Effect on heart ischemia | 15 |
| Inhibition of PGE2 | 16 |
| Inhibition of IκB kinase (IKK) | 17 |
| Mitogenesis | 18 |
| Fibrinolysis | 19 |

EETs are metabolized by various pathways, however the main pathway of their metabolism is through conversion to the corresponding 1,2 diols (dihydroxyeicosatrienoates, DHETs), has a less bioactive molecular structure that is characterized by a pro-inflammatory action.

The enzyme that carries out this reaction is the soluble epoxide hydrolase (sEH), and its inhibition could stabilize EET levels with expected beneficial biological effects (Figure 1).

The purposes of this review are: 1) to report the current status of preclinical studies on drugs inhibiting the soluble epoxide hydrolases (sEH) enzyme; 2) to evaluate the use of these novel active ingredients in veterinary medicine so that they can be used in the near future, thus increasing the veterinary drug inventory.

ARACHIDONIC ACID CASCADE AND CYP450 PATHWAY

Figure 1 shows the arachidonic acid cascade and its metabolism to eicosanoid mediators via three pathways, namely the COX, LOX and CYP450 pathways. CYP enzymes, which are mainly expressed in the liver, gut and kidney, are responsible for the metabolism of xenobiotics and many pharmaceu-

tics, but they also utilize endogenous compounds as substrates, such as cholesterol and fatty acids (6). Arachidonic acid, as shown the Figure 2, is not the only endogenous CYP substrate. CYP enzymes are also able to generate epoxides from n-6 fatty acids, linoleic acid and n-3 fatty acids, eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). Biological activity has been attributed to almost all of these CYP derivatives, however, the specific enzymes involved in the conversion of linoleic acid, EPA, and DHA are less well studied than those that metabolize AA (6). Via unique mechanisms, CYP450 metabolises EETs by incorporating them into phospholipids, chain shortening, chain elongation, hydroxylation and other pathways, however, the dominant pathway is the hydration of the epoxides to the corresponding 1,2-diols by soluble epoxide hydrolases (sEH) (20). sEH is a member of the epoxide hydrolases class which, in turn belongs to a sub-category of a broad group of hydrolytic enzymes that include esterases, proteases, dehalogenases and lipases (21).

In mammalian species, there are at least five epoxide hydrolase forms characterized by two different domains: N-terminal and C-terminal. The biological role of these domains is not yet well known but the C-terminal domain is an α/β hydrolase fold structure and is responsible for the epoxide hydrolase activity that catalyzes the hydration of chemically reactive epoxides to their corresponding diol products (21).

In conclusion, EETs have an important role especially in vascular, renal, and cardiac systems and modulated gene expression. They also induce vasorelaxation which likewise, has anti-inflammatory effects (22), but they are quickly metabolize by the sEH enzyme into corresponding less bioactive diols (20), reducing the beneficial effects of EETs. Therefore, the addition of sEH could be an efficient way to increase EET levels and to maintain their beneficial effects.

THE BIOLOGICAL EFFECTS OF SEHIS

As mentioned above, increasing EETs by sEHIs maintains their beneficial autocrine and paracrine effects. A number of studies concerning sEHIs have been carried out in different animal models (Table 2) to confirm this effect. sEHIs could be useful in the treatment of hypertension, atherosclerosis, pulmonary diseases, inflammation, and pain. However, the most studied and attractive targets are related to the treatment of inflammation, pain and cardiovascular diseases.

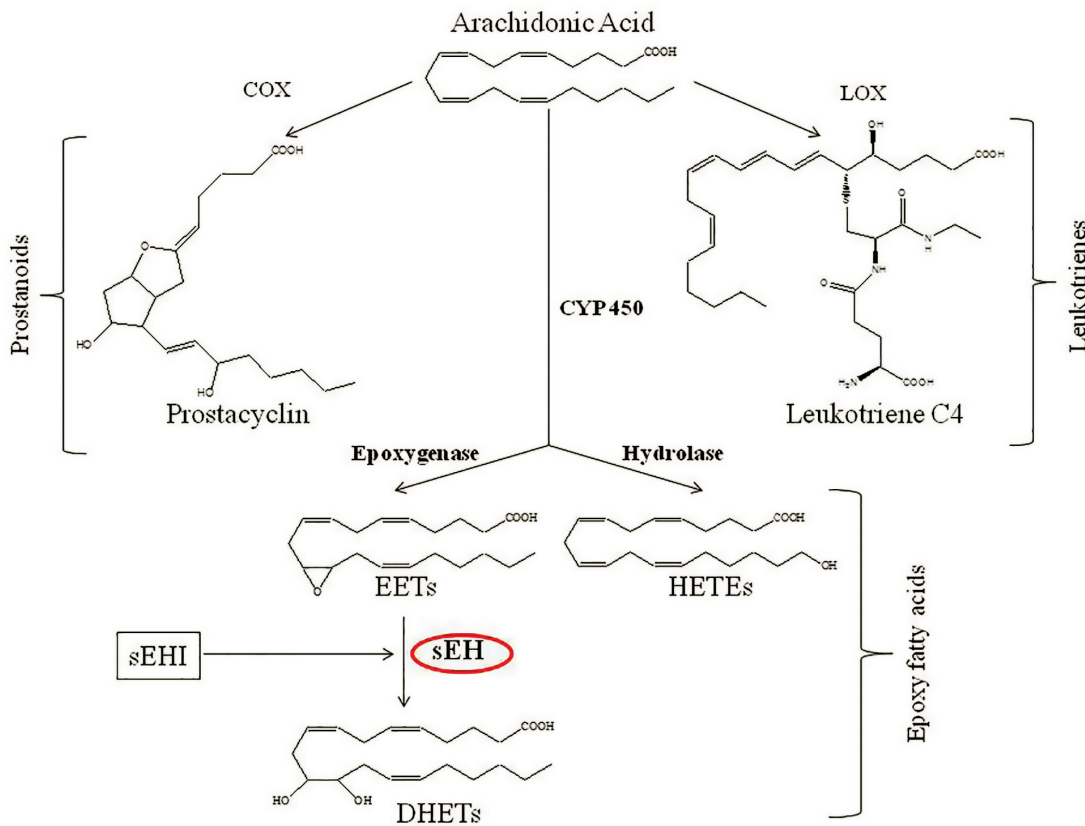


Figure 1: Arachidonic acid cascade and its major pathways metabolism. Arachidonic acid is metabolized by cyclooxygenase (COX) and lipoxygenase (LOX) enzymes into predominantly pro-inflammatory metabolites which are prostanoids and leukotrienes, respectively. The third pathway involves the cytochrome P450 enzymes that metabolize arachidonic acid into anti-inflammatory metabolites epoxy eicosatrienoic acid (EETs). EETs are rapidly metabolized by the epoxide hydrolase (sEH) to their corresponding diols dihydroxyeicosatrienoic acids (DHETs). sEH inhibitors (sEHI) block this degradation and stabilize EET levels while reducing DHETs.

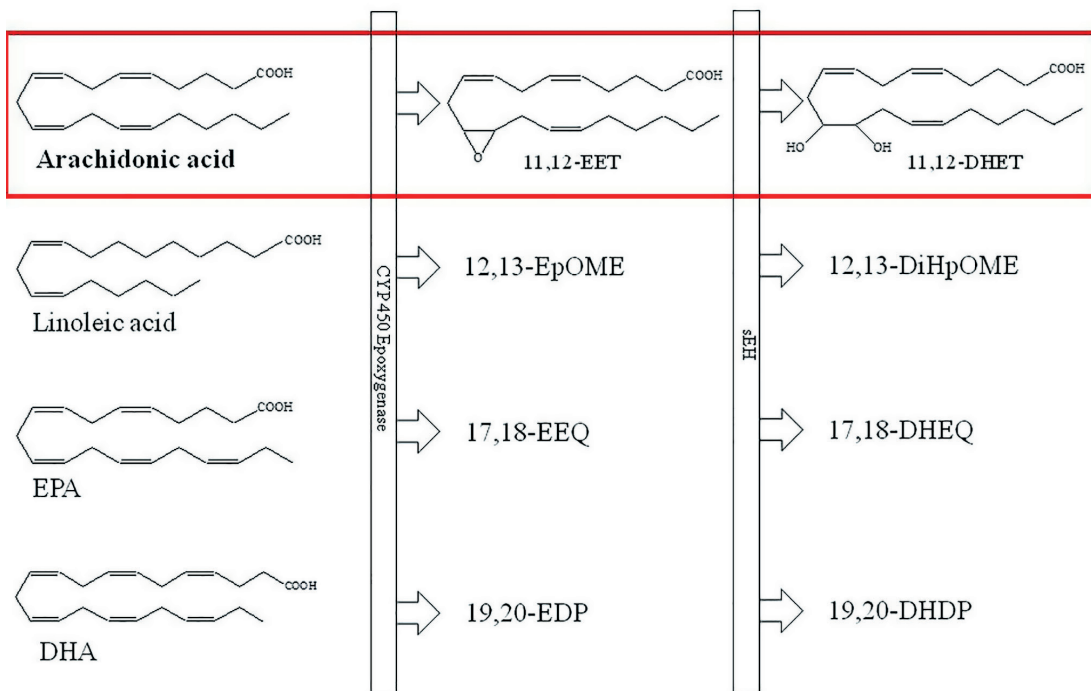


Figure 2: Endogenous CYP substrate arachidonic acid, linoleic acid, eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), their epoxydation by converting into epoxyeicosatrienoic acid (EET), epoxyoctadecenoic acid (EpOME), epoxyeicosatetraenoic acid (EEQ), epoxydocosapentaenoic acid (EDP), and metabolism of epoxides generated to the corresponding diols, by the epoxide hydrolase (sEH), dihydroxyeicosatrienoic acid (DHET), dihydroxyoctadecenoic acid (DiHOME), dihydroxyeicosatetraenoic acid (DHEQ) and dihydroxydocosapentaenoic acid (DHDP).

Table 2: Various pharmacological effects of sEHIs

| Pharmacological Effects | Species | References |
|-----------------------------------|---------|------------|
| Anti-hypertensive effect | Mouse | 26 |
| | Rat | 27, 28 |
| Myocardial protective effect | Mouse | 46, 47 |
| Anti-atherosclerosis effect | Mouse | 48 |
| Pulmonary vasoconstrictive effect | Mouse | 33 |
| Renal vasodilatory effect | Rat | 49 |
| Anti-inflammatory effect | Mouse | 35, 38 |
| | Rat | 39, 43 |
| Analgesic effect | Rat | 43, 50 |
| | Horse | 45 |

Several sEHIs have been widely used and evaluated for their biological effects in animal models. The first sEHIs discovered were substituted chalcone oxides: these demonstrated low efficacies *in vivo* (23). When the newer urea- and carbamate-based compounds were discovered, they showed a better metabolic stability and safety profile than their predecessors (24). However, none of these compounds have been launched on the market as yet, as a thorough evaluation of their therapeutic effectiveness and safety profile is lacking.

Cardiovascular effects

Vasodilation is a one of the major biological effects of the EETs (25). A number of investigators have demonstrated that sEHIs could be used to improve hypertension. These active ingredients have been shown to reduce hypertension in many animal models, with an efficacy similar to angiotensin II (26), deoxycorticosterone (27), salt and high fat diet (28). It has been reported that sEHIs reduce pulmonary vascular remodeling and delay pulmonary hypertension in monocrotaline induced pulmonary hypertension in rats (25). Imig *et al.* (29) demonstrated that the sEHI drug NCND (N-cyclohexyl-N-dodecyl urea) reduced arterial blood pressure in angiotensin II hypertensive animals. Moreover, other works showed protective effects of sEHIs against cardiovascular diseases. It has been reported that EETs reduce adverse effects of stress on mitochondrial potassium channels (30). Wang *et al.* (31) suggested that sEHIs showed a potential therapeutic effect in the treatment of atherosclerosis. The reduction of low-density lipoprotein and elevation of high density lipoprotein cholesterol were correlated with the anti-atherosclerotic effects of sEHI. In wild-type mice, sEHI

(AUDA-BE, (12-(3-adamantan-1-yl-ureido)-dodecanoic acid butyl ester)) reduced infarct size after regional myocardial ischemia-reperfusion injury *in vivo* (32) while Xu *et al.* (33), showed that sEHIs reversed cardiac hypertrophy using a murine model of pressure induced cardiac hypertrophy.

Anti-inflammatory effect

sEHIs appear to exert their anti-inflammatory effect through stabilization of EET levels. A number of investigators have demonstrated that EETs reduce inflammation. Node *et al.* (17) demonstrated that physiological concentrations of EETs or overexpression of CYP2J2 decreased cytokine-induced endothelial cell adhesion molecule expression, and EETs prevented leukocyte adhesion to the vascular wall by a mechanism involving inhibition of transcription factor NF- κ B and I κ B kinase (IKK). NF- κ B plays a key role in cytokine mediated inflammation and could be inactive while bound to I κ B. Thus, sEHIs indirectly maintain NF- κ B in the inactive state correlated with inhibition of IKK. Furthermore, inhibition of sEH enzymes led to increased anti-inflammatory properties related to regulation of cytokines (17).

Several studies have evaluated the anti-inflammatory effect of sEHIs in different inflammatory disease models. The endotoxin-induced model is a common form of the septicemic model (34); lipopolysaccharide (LPS), also known as endotoxin, is the primary Gram-negative bacteria surface antigen which causes a number of pathophysiological changes associated with eliciting immunologic responses including leukocyte activation, cytokine production and enhanced pro-inflammatory gene expression. In the LPS induced inflammation model in mice, the sEHI (AUDA-BE, (2-(3-adamantan-1-yl-ureido)-dodecanoic acid butyl ester)) decreased the production of nitric oxide metabolites and pro-inflammatory cytokines and prevented mortality (35). In another study, the sEHI (t-AUCB) significantly reduced plasma levels of pro-inflammatory cytokines such as TNF- α and IL-6 at 24 h after treatment in the LPS-treated murine model (36).

As mentioned above, EETs are converted to DHETs corresponding diols, and the blockage of this conversion by sEHIs has an important role in reducing inflammation. LPS, in particular increases the conversion into diols, decreasing the ratio of epoxides to diols. Blood epoxide and diol levels in normal animals treated with sEHIs are lower than those in inflammatory animals (37). This was clearly shown

by Liu *et al.* (38) in LPS-treated mice where sEHIs such as AUDA-BE, significantly reduced the production of diols and increased the ratio of epoxides to diols. Thus, sEHIs have been shown to have therapeutic efficacy in the treatment of endotoxin induced inflammation.

Moreover, in tobacco smoke-exposed rats, the sEHIs (AUDA-nBE, AUDA n-butyl ester) facilitated a decrease in bronchoalveolar inflammatory cells, including significant reductions in alveolar macrophages, neutrophils, and lymphocytes (39). Additionally, co-administration with EETs further reduced the number of bronchoalveolar inflammatory cells (39).

In summary, the anti-inflammatory effects of sEHIs have been shown to result through multiple pathways. sEHIs reduce the production of cytokines and pro-inflammatory lipid mediators. Stabilizing EETs by sEHIs led to down regulation of other enzymes such as COX-2 and 5-LOX in the AA cascade. In addition, the co-administration of NSAIDs and sEHIs produced an antinociceptive effect in an inflammatory pain model. Indeed a synergistic action in reducing predominantly inflammatory eicosanoids like prostaglandin PGE2 has been shown (37). Hence, it was speculated that COX inhibitors can increase EET levels and that stabilized EETs can improve anti-inflammatory effects (40). This is in agreement with results from the inflammatory rat model, where co-administration of a sEHI with a low dose of celecoxib (a COX-2 selective inhibitor) was highly effective against inflammation (41). Moreover, co-administration of sEHIs and COX inhibitors reduced the side effects of the COX inhibitor, improving their safety. Thus, the sEHIs should allow reducing the dose of COX inhibitors required for the treatment of inflammation.

Inflammatory pain

EETs dramatically reduce PGE2 levels, a cytokine with a central role in inflammation and pain, therefore, sEHIs could be used in cases of inflammatory pain to reduce production of painful mediators of inflammation. This is in line with the analgesic effect showed in animal models.

sEHI showed similar efficacy and a 1000 fold increase in potency compared to morphine in inflammatory pain

models (42). According to Inceoglu *et al.* (43), topical application of sEHIs effectively attenuates thermal hyperalgesia and mechanical allodynia in LPS-treated rats. Moreover, co-administration of EETs with a sEHI showed an additive increase in anti-hyperalgesia and sEHIs were demonstrated as acting in both peripheral and central nerves systems (44).

Veterinary applications

In the field of veterinary medicine, the first application of sEHI was conducted in 2013. This study tested if the sEHI (t-TUCB, trans-4-{4-[3-(4-trifluoromethoxy-phenyl)-ureido]-cyclohexyloxy}-benzoic acid) might reduce severe inflammatory pain in a horse affected by laminitis (45). The patient in this study was a horse treated for laminitis for a 7-day period using different NSAIDs and gabapentin. Treatments with these classical drugs were not effective and euthanasia was being considered for humane reasons. On day 8, a sEHI was added to the treatment protocol. After the first dose, the horse began to walk spontaneously, had a good appetite with remarkable reduction in pain scores and no side effects were reported (45).

In conclusion, the studies presented in this review are quite persuasive in demonstrating that sEHIs may have potential applications in the treatment of several diseases. Moreover, a recent study has also demonstrated effectiveness of these drugs in veterinary medicine (45). Hopefully, this is the first of many sets of data to be generated regarding the successful treatment of pain in animals. These new active ingredients could be particularly applicable in animals sensitive to the common anti-inflammatory drugs.

CONFLICT OF INTERESTS

None of the authors has any financial or personal relationship that could inappropriately influence the content of the paper.

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REFERENCES

- Girogi, M.: Veterinary Pharmacology: Is it Still Pharmacology's Cinderella? *Clin. Exp. Pharmacol.* 2: 2, 2012.
- Kim, T.W. and Girogi, M.: A brief overview of the coxib drugs in the veterinary field. *Am. J. Anim. Vet. Sci.* 8: 89-97, 2013.
- Lavy, E., Prise, U., Soldani, G., Neri, D., Brandriss, N., Bar Chaim, A. and Girogi, M.: Pharmacokinetics of methylphenidate after oral administration of immediate and sustained-release preparations in Beagle dogs. *Vet. J.* 189: 336-340, 2011.
- Giorgi, M., Saccomanni, G., Del Carlo, S., Manera, C. and Lavy, E.: Pharmacokinetics of intravenous and intramuscular parecoxib in healthy Beagles. *Vet. J.* 193: 246-250, 2012.
- Giorgi, M., Mills, P.C., Tayari, H., Rota, S., Breggi G. and Briganti, A.: Plasma concentrations of tapentadol and clinical evaluations of a combination of tapentadol plus sevoflurane for surgical anaesthesia and analgesia in rabbits (*Oryctolagus cuniculus*) undergoing orchietomy. *Isr. J. Vet. Med.* 68: 141-148, 2013.
- Fleming, I.: The cytochrome P450 pathway in angiogenesis and endothelial cell biology. *Cancer. Metast. Rev.* 30: 541-555, 2011.
- Wagner, K., Inceoglu, B. and Hammock, B.D.: Soluble epoxide hydrolase inhibition epoxygenated fatty acids and nociception. *Prostag. Oth. Lipid. M.* 96: 76-83, 2011.
- Lee, H.C., Lu, T., Weintraub, N.L., VanRollins, M., Spector, A.A. and Shibata, E.F.: Effects of epoxyeicosatrienoic acids on sodium channels in isolated rat ventricular myocytes. *J. Physiol.* 519: 153-168, 1999.
- Sakairi, Y., Jacobson, H.R., Noland, T.D., Capdevila, J.H., Falck, J.R. and Breyer, M.D.: 5,6-EET inhibits ion transport in collecting duct by stimulating endogenous prostaglandin synthesis. *Am. J. Physiol-renal.* 268: 931-939, 1995.
- Lu, T., Vanrollins, M. and Lee, H.C.: Stereospecific activation of cardiac ATP-sensitive K⁺ channels by epoxyeicosatrienoic acids: a structural determinant study. *Mol. Pharmacol.* 62: 1076-1083, 2002.
- Vriens, J., Owsianik, G., Fisslthaler, B., Suzuki, M., Janssens, A., Voets, T., Morisseau, C., Hammock, B.D., Fleming, I., Busse, R. and Nilius, B.: Modulation of Ca²⁺ permeable cation channel TRPV4 by cytochrome P450 epoxygenases in vascular endothelium. *Circ. Res.* 97: 908-915, 2005.
- Graier, W.F., Simecek, S. and Sturek, M.: Cytochrome P450 mono-oxygenase-regulated signalling of Ca²⁺ entry in human and bovine endothelial cells. *J. Physiol.* 15: 259-274, 1995.
- Benoit, C., Renaudon, B., Salvail, D. and Rousseau, E.: EETs relax airway smooth muscle via an EpDHF effect: BF(a) channel activation and hyperpolarization. *Am. J. Physiol. Lung. Cell. Mol. Physiol.* 280: 965-973, 2001.
- Moffat, M.P., Ward, C.A., Bend, J.R., Mock, T., Farhangkhome, P. and Karmazyn, M.: Effects of epoxyeicosatrienoic acids on isolated hearts and ventricular myocytes. *Am. J. Physiol-Heart. C.* 26: 1154-1160, 1993.
- Wu, S., Chen, W., Murphy, E., Gabel, S., Tomer, K.B., Foley, J., Steenbergen, C., Falck, J.R., Moomaw, C.R. and Zeldin, D.C.: Molecular cloning, expression, and functional significance of a cytochrome P450 highly expressed in rat heart myocytes. *J. Biol. Chem.* 272: 12551-12559, 1997.
- Fang, X., Moore, S.A., Stoll, L.L., Rich, G., Kaduce, T., Weintraub, N. and Spector, A.A.: 14, 15-Epoxyeicosatrienoic acid inhibits prostaglandin E2 production in vascular smooth muscle cells. *Am. J. Physiol-Heart. Circ. Physiol.* 275: 2113-2121, 1998.
- Node, K., Huo, Y., Ruan, X., Yang, B., Spiecker, M., Ley, K., Zeldin, D.C. and Liao, J.K.: Anti-inflammatory properties of cytochrome P450 epoxygenase-derived eicosanoids. *Science.* 20: 1276-1279, 1999.
- Chen, J.K., Capdevila, J. and Harris, R.C.: Overexpression of C-terminal Src kinase blocks 14, 15-epoxyeicosatrienoic acid-induced tyrosine phosphorylation and mitogenesis. *J. Biol. Chem.* 275: 13789-13792, 2000.
- Node, K., Ruan, X.L., Dai, J., Yang, S.X., Graham, L., Zeldin, D.C. and Liao, J.K.: Activation of G α s mediates induction of tissue-type plasminogen activator gene transcription by epoxyeicosatrienoic acids. *J. Biol. Chem.* 276: 15983-15989, 2001.
- Shen, H.C. and Hammock, B.D.: Discovery of inhibitors of soluble epoxide hydrolase: a target with multiple potential therapeutic indications. *J. Med. Chem.* 55: 1789-1808, 2012.
- Fretland, A.J. and Omiecinski, C.J.: Epoxide hydrolases: biochemistry and molecular biology. *Chem. Biol. Interact.* 129: 41-59, 2000.
- Spector, A.A., Fang, X., Snyder, G.D. and Weintraub, N.L.: Epoxyeicosatrienoic acids (EETs): metabolism and biochemical function. *Prog. Lipid Res.* 43: 55-90, 2004.
- Morisseau, C. and Hammock, B.D.: Epoxide hydrolases: Mechanisms, inhibitor designs, and biological roles. *Annu. Rev. Pharmacol. Toxicol.* 45: 311-333, 2005.
- Morisseau, C., Goodrow, M.H., Dowdy, D., Zheng, J., Greene, J.F., Sanborn, J.R. and Hammock, B.D.: Potent urea and carbamate inhibitors of soluble epoxide hydrolases. *Proc. Natl. Acad. Sci.* 96: 8849-8854, 1999.
- Spector, A.A. and Norris, A.W.: Action of epoxyeicosatrienoic acids on cellular function. *Am. J. Physiol. Cell. Physiol.* 292: C996-1012, 2007.
- Hercule, H.C., Schunck, W.H., Gross, V., Seringer, J., Leung, F.P., Weldon, S.M., da Costa Goncalves, A.C., Huang, Y., Luft, F.C. and Gollasch, M.: Interaction between P450 eicosanoids and nitric oxide in the control of arterial tone in mice. *Arterioscler. Thromb. Vasc. Biol.* 29: 54-60, 2009.
- Loch, D., Hoey, A., Morisseau, C., Hammock, B.D. and Brown, L.: Prevention of hypertension in DOCA-salt rats by an inhibitor of soluble epoxide hydrolase. *Cell. Biochem. Biophys.* 47: 87-98, 2007.
- Huang, H., Morisseau, C., Wang, J., Yang, T., Falck, J.R., Hammock, B.D. and Wang, M.H.: Increasing or stabilizing renal epoxyeicosatrienoic acid production attenuates abnormal renal function and hypertension in obese rats. *Am. J. Physiol. Renal. Physiol.* 293: 342-349, 2007.
- Imig, J.D., Zhao, X., Capdevila, J.H., Morisseau, C. and Hammock, B.D.: Soluble epoxide hydrolase inhibition lowers arterial blood pressure in angiotensin II hypertension. *Hypertension.* 39: 690-694, 2002.
- Seubert, J.M., Sinal, C.J., Graves, J., DeGraff, L.M., Bradbury, J.A., Lee, C.R., Goralski, K., Carey, M.A., Luria, A., Newman, J.W., Hammock, B.D., Falck, J.R., Roberts, H., Rockman, H.A., Murphy, E. and Zeldin, D.C.: Role of soluble epoxide hydrolase

- in postischemic recovery of heart contractile function. *Circ. Res.* 99: 442-450, 2006.
31. Wang, Y.J., Ulu, A., Zhang, L.N. and Hammock, B.D.: Soluble Epoxide Hydrolase in Atherosclerosis. *Curr. Atheroscler. Rep.* 12: 174-183, 2010.
 32. Motoki, A., Merkel, M.J. and Packwood, W.H.: Soluble epoxide hydrolase inhibition and gene deletion are protective against myocardial ischemia-reperfusion injury in vivo. *Am. J. Physiol. Heart Circ. Physiol.* 295: 2128-2134, 2008.
 33. Xu, D., Li, N., He, Y., Timofeyev, V., Lu, L., Tsai, H.J., Kim, I.H., Tuteja, D., Mateo, R.K.P., Singapuri, A., Davis, B.B., Low, R., Hammock, B.D. and Chiamvimonvat, N.: Prevention and reversal of cardiac hypertrophy by soluble epoxide hydrolase inhibitors. *Proc. Natl. Acad. Sci. USA.* 103: 18733-18738, 2006.
 34. Kanaan, S.A., Saade, N.E., Haddad, J.J., Abdelnoor, A.M., Atweh, S.F., Jabbur, S.J. and Saileh-Garabedian, B.: Endotoxin-induced local inflammation and hyperalgesia in rats and mice: a new model for inflammatory pain. *Pain.* 66: 373-379, 1996.
 35. Schmelzer, K.R., Kubala, L., Newman, J.W., Kim, I.H., Eiserich, J.P. and Hammock, B.D.: Soluble epoxide hydrolase is a therapeutic target for acute inflammation. *Proc. Natl. Acad. Sci. USA.* 102: 9772-9777, 2005.
 36. Liu, J.Y., Yang, J., Inceoglu, B., Qiu, H., Ulu, A., Hwang, S.H., Chiamvimonvat, N. and Hammock, B.D.: Inhibition of soluble epoxide hydrolase enhances the anti-inflammatory effects of aspirin and 5-lipoxygenase activation protein inhibitor in a murine model. *Biochem. Pharmacol.* 79: 880-887, 2010.
 37. Schmelzer, K.R., Inceoglu, B., Kubala, L., Kim, I.H., Jinks, S.L. and Eiserich, J.P.: Enhancement of antinociception by coadministration of nonsteroidal anti-inflammatory drugs and soluble epoxide hydrolase inhibitors. *Proc. Natl. Acad. Sci. USA.* 103: 13646-13651, 2006.
 38. Liu, J.Y., Tsai, H.J., Hwang, S.H., Jones, P.D., Morisseau, C. and Hammock, B.D.: Pharmacokinetic optimization of four soluble epoxide hydrolase inhibitors for use in a murine model of inflammation. *Brit. J. Pharmacol.* 156: 284-296, 2009.
 39. Smith, K.R., Pinkerton, K.E., Watanabe, T., Pedersen, T.L., Ma, S.J. and Hammock, B.D.: Attenuation of tobacco smoke-induced lung inflammation by treatment with a soluble epoxide hydrolase inhibitor. *Proc. Natl. Acad. Sci. USA.* 102: 2186-2191, 2005.
 40. Inceoglu, B., Schmelzer, K.R., Morisseau, C., Jinks, S.L. and Hammock, B.D.: Soluble epoxide hydrolase inhibition reveals novel biological functions of epoxyeicosatrienoic acids (EETs). *Prostag. Oth. Lipid. M.* 82: 42-49, 2007.
 41. Hwang, S.H., Wagner, K. and Morisseau, C.: Synthesis and structure-activity relationship studies of urea-containing pyrazoles as dual inhibitors of cyclooxygenase-2 and soluble epoxide hydrolase. *J. Med. Chem.* 54: 3037-3050, 2011.
 42. Rose, T.E., Morisseau, C., Liu, J.Y., Inceoglu, B., Jones, P.D., Sanborn, J.R. and Hammock, B.D.: 1-Aryl-3-(1-acylpiperidin-4-yl) urea inhibitors of human and murine soluble epoxide hydrolase: structure-activity relationships, pharmacokinetics, and reduction of inflammatory pain. *J. Med. Chem.* 53: 7067-7075, 2010.
 43. Inceoglu, B., Jinks, S.L., Schmelzer, K.R., Waite, T., Kim, I.H. and Hammock, B.D.: Inhibition of soluble epoxide hydrolase reduces LPS-induced thermal hyperalgesia and mechanical allodynia in a rat model of inflammatory pain. *Life Sci.* 79: 2311-2319, 2006.
 44. Wagner, K., Inceoglu, B., Gill, S.S. and Hammock, B.D.: Epoxy-generated fatty acids and soluble epoxide hydrolase inhibition: novel mediators of pain reduction. *J. Agric. Food Chem.* 59: 2816-2824, 2011.
 45. Guedes, A.G.P., Morisseau, C., Sole, A., Soares, J.H.N., Ulu, A., Dong, H. and Hammock, B.D.: Use of a soluble epoxide hydrolase inhibitor as an adjunctive analgesic in a horse with laminitis. *Vet. Anaesth. Analg.* 40: 440-448, 2013.
 46. Zhang, L.N., Vincelette, J., Cheng, Y., Mehra, U., Chen, D., Anandan, S.K., Gless, R., Webb, H.K. and Wang, Y.X.: Inhibition of soluble epoxide hydrolase attenuated atherosclerosis, abdominal aortic aneurysm formation, and dyslipidemia. *Arterioscler. Thromb. Vasc. Biol.* 29: 1265-1270, 2009.
 47. Keseru, B., Barbosa-Sicard, E., Popp, R., Fisslthaler, B., Dietrich, A., Gudermann, T., Hammock, B.D., Falck, J.R., Weissmann, N., Busse, R. and Fleming, I.: Epoxyeicosatrienoic acids and the soluble epoxide hydrolase are determinants of pulmonary artery pressure and the acute hypoxic pulmonary vasoconstrictor response. *Faseb. J.* 22: 4306-4315, 2008.
 48. Li, N., Liu, J.Y., Timofeyev, V., Qiu, H., Hwang, S.H., Tuteja, D., Lu, L., Yang, J., Mochida, H., Low, R., Hammock, B.D. and Chiamvimonvat, N.: Beneficial effects of soluble epoxide hydrolase inhibitors in myocardial infarction model: insight gained using metabolomics approaches. *J. Mol. Cell. Cardiol.* 47: 835-845, 2009.
 49. Carroll, M.A., Doumad, A.B., Li, J., Cheng, M.K., Falck, J.R. and McGiff, J.C.: Adenosine2A receptor vasodilation of rat preglomerular microvessels is mediated by EETs that activate the cAMP/PKA pathway. *Am. J. Physiol. Renal. Physiol.* 291: 155-161, 2006.
 50. Inceoglu, B., Jinks, S.L., Ulu, A., Hegedus, C.M., Georgi, K., Schmelzer, K.R., Wagner, K., Jones, P.D., Morisseau, C. and Hammock, B.D.: Soluble epoxide hydrolase and epoxyeicosatrienoic acids modulate two distinct analgesic pathways. *Proc. Natl. Acad. Sci. USA.* 105: 18901-18906, 2008.